

Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Disease



journal homepage: www.elsevier.com/locate/apjtd

Document heading doi: 10.1016/S2222-1808(14)60431-X

© 2014 by the Asian Pacific Journal of Tropical Disease. All rights reserved.

## Phytochemical screening and antioxidant activity of methanolic extract of selected wild edible Nigerian mushrooms

Hamzah Rabiat Unekwu<sup>°</sup>, Jigam Ali Audu, Makun Hussaini Makun, Egwim Evans Chidi

Department of Biochemistry, School of Natural and Applied Sciences Federal University of Technology, P. M. B. 65, Minna, Niger State, Nigeria

#### PEER REVIEW

#### Peer reviewer

Dr. Abdullahi Mann, Department of Chemistry, Federal University of Technology, P. M. B. 65, Minna, Niger State, Nigeria. Tel: +2348034295656:

E-mail: abdumann@yahoo.com

#### **Comments**

This is a good study in which the authors phytochemically investigate as well as demonstrate the antioxidant activity of eight wild edible Nigerian mushrooms. DPPH free radical was used to evaluate the scavenging ability of the mushroom extracts and their reducing power were also assessed. Details on Page S156

#### ABSTRACT

**Objective:** To elucidate the phytochemical content and antioxidant activity of selected wild edible Nigerian mushroom species.

**Methods:** Phytochemical screening was carried out using standard methods while 1,1–Diphenyl picryl hydrazyl (DPPH) radical and reductive power assays were used to evaluate the *in vitro* antioxidant properties of the selected edible Nigerian mushroom species.

**Results:** The result obtained revealed the presence of alkaloids, cardiac glycosides, saponins, flavonoids, terpenes, steroids, tannins and phenols in the selected mushrooms extracts. The extract of *Pleutorus ostearus* showed a significantly (*P*<0.05) higher total phenol and flavonoid content of (248.80±7.63) mg/g and (42.63±0.63) mg/g respectively compared to other mushroom extracts. *Cantherale cibarus* had the most significant (*P*<0.05) amount of alkaloids [(135.57±0.27) mg/g] and saponins [(150.41±0.50) mg/g] when compared to other extracts while the tannin content [(170.56±0.74)] mg/g was highest in the mushroom *Temitomyces robustus*. All mushroom extracts scavenged DPPH radical in a dose dependent manner. However, *Lactarus deliciousus* had the highest DPPH scavenging activity compared to the other mushroom extracts. *Pleutorus ostearus* and *Lactarus deliciousus* had better reductive power than other mushroom extracts concentrations used.

**Conclusions:** The mushroom species analysed have been shown to be good sources of antioxidants and other phytoconstituents, thus it can be used in the management of oxidative stress induced diseases.

#### KEYWORDS

Phytochemicals, Antioxidants, Mushrooms, 1,1- Diphenyl picryl hydrazyl, Reductive power

#### **1. Introduction**

Free radicals are constantly formed in the human body during energy production, in the mitochondrial electron transport chain, phagocytosis, arachidonic acid metabolism, ovulation, fertilization and in xenobiotic metabolism<sup>[1]</sup> and from external sources such as food, drugs, smoke and other pollutants in the environment<sup>[2]</sup>. Living organisms are endowed with endogenous and exogenous antioxidant defense systems capable of countering the adverse reactions of free radicals<sup>[3]</sup>. The generation of free radicals in the body beyond its antioxidant capacity actually leads to oxidative stress and this has been implicated in the etiology of a

E-mail: rabiune@yahoo.com

number of disorders<sup>[4]</sup>. As a result, of this much attention is being focused on the use of antioxidants to inhibit and protect damage due to free radicals and reactive oxygen species. Synthetic antioxidants such as butylated hydroxyanisole, tert-butylated hydroxyquinone and butylated hydroxytoluene are radical scavengers but are usually associated with adverse side effects<sup>[5]</sup>. Neutralization of radical damage by naturally occurring antioxidants from several sources either as food supplements or drugs is becoming one of the most acceptable modes of modern therapy<sup>[6]</sup>.

Mushrooms have continued to generate a lot of interest particularly in their consumption as food[7], in the cure

Article history: Received 15 Nov 2013 Received in revised form 26 Nov, 2nd revised form 4 Dec, 3rd revised form 12 Dec 2013 Accepted 28 Dec 2013 Available online 28 Jan 2014

<sup>\*</sup>Corresponding author: Hamzah Rabiat Unekwu, Department of Biochemistry, School of Natural and Applied Sciences, Federal University of Technology, P. M. B. 65, Minna, Niger State, Nigeria.

Foundation Project: Supported by the University Board of Research of Federal University of Technology Minna, Niger State, Nigeria.

of diseases<sup>[8]</sup>, in bioremediation and as important items of commerce all over the world due to their nutritional, antioxidant and therapeutic values<sup>[9,10]</sup>. They may then be utilized to be amongst the useful candidates in the search for bioactive compounds with radical scavenging activity<sup>[11,12]</sup>. Although there are many studies on nutrients compositions of different mushroom species, only few studies have been carried out on the antioxidant activity in wild edible species<sup>[12,13]</sup>. Therefore this study was carried out to elucidate the phytochemical composition and *in vitro* antioxidant activities of methanolic extract of the selected Nigerian wild edible mushrooms.

#### 2. Materials and methods

#### 2.1. Collection of samples

Eight indigenous wild edible Nigerian mushrooms including Cantharelle cibarius (C. cibarius), Termitomyces robustus (T. robustus), Termitomyces manniformis (T. manniformis), Pleurotus ostreatus (P. ostreatus), Pleurotus pulmonarius (P. pulmonarius), Auricularia cularia (A. cularia), Hericium erinaceus (H. erinaceus), Lactarus deliciousus (L. deliciousus) were collected from logs of wood, palm logs and humus soil from different locations in Nigeria. They were identified by a Taxonomist, Prof. Onyekwere S. C. of Applied Biology Department, Ebonyi State University Abakaliki, Nigeria.

#### 2.2. Sample preparation and extraction

Mushrooms were destalked and air dried at room temperature with adequate ventilation and pulverized using a blender. The pulverized samples were extracted with methanol by reflux. Exactly 50 g of the powdered samples were weighed into 400 mL of methanol in a reflux flask and refluxed for 2 h. The extracts were filtered hot using a muslin cloth and subsequently evaporated using a rotary evaporator. The semi-dry extracts were weighed, placed in sterile sample bottles and stored in a refrigerator until required for use.

#### 2.3. Qualitative phytochemical screening

Table 1

The extracts were screened for phytochemical properties

Phytochemical constituents of selected wild edible Nigerian mushroom species.

using standard methods<sup>[14]</sup>.

### 2.4. Quantitative determination of the phytochemical constituents in samples

Aluminum chloride colorimetric method was used for flavonoid determination<sup>[15]</sup> while total phenol content of the extracts was determined using the method reported by Singleton *et al*<sup>[16]</sup>.

The method of Oloyed<sup>[17]</sup> was used to determine the amount of alkaloids and saponins in the mushroom extracts while tannin content was quantified with the method described by AOAC<sup>[18]</sup>.

#### 2.5. In vitro antioxidant determinations

Ability of the extracts to scavenge 1,1-diphenyl-2 picrylhydrazyl (DPPH) free radical was evaluated as described by Gyamfi *et al.*<sup>[19]</sup> and the reducing power of the extracts was determined by assessing the ability of the extracts to reduce FeCl<sub>3</sub> solution as described by Oyaizu<sup>[20]</sup>.

#### 2.6. Statistical analysis

All values were expressed as mean±SEM. The SPSS program (version 16.0 SPSS Inc., Chicago, IL, USA) was used for the analysis of variance followed by the new Duncan multiple test.

#### 3. Results

#### 3.1. Qualitative phytochemical screening

Phytochemical sreening result revealed the presence of alkaloids, cardiac glycosides, saponins, flavonoids, terpenes, steroids, tannins and phenolics in the selected mushroom extracts in varying proportions (Table 1). Phlobatannins was absent in all mushrooms except *T. manniformis* and *P. ostreatus* while anthraquinone was present in all except *H. erinaceus*, *A. cularia* and *P. ostreatus*.

#### 3.2. Quantitative phytochemical analysis

The quantitative phytochemical content determination of the methanolic extract of the selected mushroom result

				*						
Mushrooms	Alkaloids	Anthraquinones	Cardiac glycosides	Flavonoids	Phenols	Phlobatannins	Saponins	Tannin	Terpenes	Steroids
C. cibarius	+ + +	+	+ +	+ +	+	-	+	+	+ + +	+ + +
T. robustus	+ +	+ + +	+ + +	+ +	+ +	-	+ +	++	+ + +	+ +
T. manniformis	+ +	+	+ + +	+ +	+ +	+	+ + +	+ +	+ + +	+ + +
P. pulmonarius	+	+ +	+ +	+ +	+ +	-	+ +	+	+ +	+ +
P. ostreatus	+ +	-	+ + +	+ +	+ +	+ +	+ +	+ +	+ +	+ +
L. deliciousus	+	+ +	+ + +	+ +	+	-	+ +	+ +	+ + +	+
A. auricula	+	-	+ + +	+	+	-	+ +	+	+ + +	+ +
H. erinaceus	+	_	+ + +	+	+	-	+	+	+	+

-: Absent, +: Faintly present, ++: Moderately present, +++: Highly present.

#### Table 2

Quantitative phytochemical contents of selected wild edible Nigerian mushroom species.

Phytochemicals	C. cibarius	T. robustus	T. manniformis	P. pulmonarius	P. ostreatus	L. deliciousus	A. auricula	H. erinaceus
Alkaloids (µg/g)	$135.57 \pm 0.27^{h}$	$85.29 \pm 0.04^{f}$	$112.51 \pm 0.04^{g}$	$46.90 \pm 0.73^{\circ}$	$81.51 \pm 0.73^{e}$	$52.99 \pm 0.07^{d}$	$16.3200 \pm 0.46^{b}$	$8.1250 \pm 0.40^{a}$
Saponins (mg/g)	$150.41 \pm 0.50^{h}$	$87.56 \pm 0.81^{f}$	$105.30 \pm 4.53^{g}$	$34.76 \pm 1.46^{\circ}$	$84.95 \pm 5.03^{e}$	$20.35 \pm 1.82^{b}$	$71.21 \pm 0.50^{d}$	$10.17 \pm 1.09^{a}$
Tannins (mg/g)	$76.29 \pm 0.74^{\circ}$	$170.56 \pm 0.74^{g}$	$169.19 \pm 0.5^{g}$	$88.91 \pm 0.16^{d}$	$146.30 \pm 0.8^{f}$	$85.73 \pm 0.8^{a}$	$66.9250 \pm 0.33^{b}$	$59.27{\pm}0.00\mathrm{b}^{\mathrm{a}}$
Flavonoids (mg/g)	$11.50 \pm 0.50^{b}$	$23.88 \pm 1.13^{cd}$	$25.66 \pm 0.74^{d}$	$22.17 \pm 0.88^{\circ}$	$42.63 \pm 0.63^{f}$	$34.58 \pm 0.93^{e}$	$6.41 \pm 0.53^{a}$	$6.75 \pm 0.38^{a}$
Phenols (mg/g)	$97.16 \pm 0.94^{a}$	$177.96 \pm 1.72^{\circ}$	$211.82 \pm 2.42^{d}$	$223.11 \pm 0.02^{e}$	$248.80 \pm 7.63^{f}$	$115.68 \pm 2.1^{b}$	$115.99 \pm 0.80^{b}$	$105.09 \pm 3.3^{a}$

Results are presented as mean±SEM. Letters represent the level of significance.

as given in Table 2 showed that alkaloid contents of the extracts ranged between  $8.125-135.57 \mu g/g$ , tannin contents of the extract ranged between  $(59.27\pm0.00)-(170.56\pm0.74) mg/g$ , saponin  $(10.17\pm1.09)-(150.41\pm0.50) mg/g$ , total phenols  $(97.16\pm0.94)-(248.80\pm7.63) mg/g$  and total flavonoid contents  $(6.41\pm0.53)-(42.63\pm0.63) mg/g$ . The extracts of *P. ostreatus* and *L. deliciousus* showed a significantly high (*P*<0.05) total flavonoid content of  $(42.63\pm0.63) mg/g$  and  $(34.58\pm0.93) mg/g$  respectively compared to all other mushroom extracts. Methanolic extract of *P. osteatus* also showed the highest total phenol content of  $(248.80\pm7.63) mg/g$  while *C. cibarius* had the most significant amount of alkaloids [(135.57\pm0.27) mg/g] and saponins [(150.41\pm0.50) mg/g] when compared to other extracts.

#### 3.3. Antioxidative properties

#### 3.3.1. DPPH radical scavenging activities

The DPPH radical scavenging activity of the selected wild edible Nigerian mushroom species is shown in Figure 1. All the mushroom extracts scavenged the DPPH radical in a dose dependent manner. However *L. deliciousus* had the highest DPPH scavenging activity compared to other mushroom extracts (Figure 1) with values ranging from 14.29% to 71.49%.



rigure 1. DPPH scavenging activity of selected wild edible Nige mushroom species.

#### 3.3.2. Reductive power

Results of the reductive power of the selected wild edible Nigerian mushroom species indicate that activities of the extracts were proportional to concentration (200–1000 mg/ mL). *P. ostreatus* and *L. deliciousus* had better reductive power than other mushroom extracts irrespective of the concentrations (Figure 2).



#### igure 2. Reductive power of selected wild cubic regenan musinoon s

#### 4. Discussion

The results obtained from the qualitative phytochemical screening of methanolic extracts of the selected wild edible Nigerian mushroom species showed the presence of tannin, saponins, cardiac glycosides, alkaloids, steroids, terpenes, phenols, and flavonoid in varying concentrations. These results agree with previous work on some selected mushrooms in Nigeria and Sudan<sup>[12,21]</sup> and conform to some a certain degree with the study on some mushrooms from Kenya<sup>[22]</sup>. The absence of anthraquinones in *H. erinaceus*, *A. cularia* and *P. ostreatus* correlates with that of previous reports in the literature<sup>[12,22]</sup>. These phytoconstituents play a significant role in the medicinal properties of many plants.

Saponins for instance comprise a large family of structurally related compounds containing a steroid or triterpernoid aglycone. They are reported to have a wide range of beneficial pharmacological properties, such as anti–inflammatory and anti–diabetic effects<sup>[23]</sup>. Thus these mushrooms can be used in the management of diabetes and inflammation related diseases.

Terpenoids have been reported to show a wide range of pharmacological benefits that include anti-malarial, antiinflammatory and anti-cancer effects among others<sup>[24,25]</sup>.

The valuable pharmacological properties of many mushrooms have also been attributed to the presence of alkaloids on the autonomic nervous system, blood vessels, respiratory system, gastrointestinal tract, uterus, and have been shown to be effective against malignant diseases, infections and malaria<sup>[26]</sup>. Phenolic compounds are antioxidants, and exhibit a wide range spectrum of medicinal properties such as anti-cancer, ant-inflammatory and diabetic effects<sup>[27,28]</sup>.

# Flavonoids are one of the most diverse group of natural compounds that have been shown to possess a broad spectrum of chemical and biological activities including radical scavenging properties, antiallergenic, antiviral, antiinflammatory, and vasodilating actions<sup>[29,30]</sup>. Thus the extracts of the studied mushrooms may be good alternatives for the treatment of diseases associated with excessive free radical generation and damage.

The high flavonoid content in *P. ostreatus* and *L. deliciousus* was found to be higher than that found in an edible mushroom  $[(2.84\pm0.12) \text{ mg/g}]$  in a recent study<sup>[31]</sup>. The phenols content also in *P. ostreatus* and *P. pulmonarius* were found to be higher than that of some wild edible mushrooms investigated in recent study<sup>[32]</sup>.

These mushrooms can therefore be harnessed in the management of oxidative stress induced diseases since phenols and flavonoid have been shown to posses various antioxidant functions.

DPPH free radical scavenging activity assay is one of the most common methods for the determination of antioxidant capacity. It relies on the reduction of methanolic DPPH solution in the presence of hydrogen donation compound (antioxidant). The resulting decolourisation upon abstraction of hydrogen from the antioxidant is stoichiometric with respect to the degree of reduction and absorbance measurement after a certain time corresponds inversely to the radical scavenging activity of the antioxidant<sup>[33]</sup>.

Although the mushroom, *P. ostreatus* specie had the highest amount of phenol [(248.80 $\pm$ 7.63) mg/g] and flavonoids [(42.63 $\pm$ 0.63) mg/g] compared to other mushrooms extracts, it did not have the highest scavenging activity, rather the mushroom *L. deliciousus* had the most appreciable DPPH scavenging activity amongst other mushrooms in a dose dependent manner.

Thus other non-phenolic and flavonoids compound may be responsible for the antioxidant properties of *L. deliciousus* while *P. ostreatus* may have exhibted its antioxidant activity through a different mechanism.

Fe (III) reduction is often used as an indicator of electron donating activity, which is an important mechanism of antioxidant action of phenolics<sup>[34]</sup>. *P. ostreatus* showed a higher reductive power than other mushroom extracts in a concentration dependent manner and this commensurate with the high phenol and flavonoid content in this mushroom. This supports earlier reports, correlating the presence of flavonoids and phenolic compounds to antioxidative actions<sup>[35,36]</sup>. Therefore, the *in vitro* antioxidant properties exhibited by this mushroom extract may be due to the presence of these antioxidant phytochemicals inherent in it and these mushrooms

#### **Conflict of interest statement**

We declare that we have no conflict of interest.

#### Acknowledgements

The authors wish to express their profound gratitude to the University Board of Research of Federal University of Technology Minna, Niger State, Nigeria for the Financial assistance awarded towards this study.

#### Comments

#### Background

Globally mushrooms are major food. Indigenous wild edible mushrooms are important to Nigerians. Therefore, it is necessary to the study its phytochemical composition and antioxidant activities.

#### Research frontiers

The present research work demonstrates *in vitro* antioxidant activity of methanolic extract of eight indigenous wild edible Nigerian mushrooms by screening them against DPPH radical and reductive power assays. *Related reports* 

#### Experimental evidence suggests that free radicals and reactive oxygen species are involved in human diseases. Plants including indigenous wild edible mushrooms produce a lot of antioxidants to control the oxidative stress induced diseases. And they can represent a source of new compounds with antioxidant activity. DPPH radical and reductive power assays are used for its evaluation.

#### Innovations and breakthroughs

Mushrooms are widely known for their culinary and medicinal properties and very often taken as food globally. In the present study, authors have demonstrated the antioxidant activity of eight indigenous wild edible Nigerian mushrooms against DPPH radical and reductive power assays.

#### **Applications**

From the literature review, it has been found that some indigenous wild edible Nigerian mushrooms are safe to humans. This scientific study supports their ethnobotanical uses particularly as antioxidant agent for the management of oxidative stress induced diseases.

#### Peer review

This is a good study in which the authors phytochemically investigate as well as demonstrate the antioxidant activity of eight wild edible Nigerian mushrooms. DPPH free radical was used to evaluate the scavenging ability of the mushroom extracts and their reducing power were also assessed.

#### References

 Halliwell B, Gutteridge J. Free radicals in biology and medicine. 4th edition, Oxford, UK: Oxford University Press; 2007.

- [2] Miller RA, Britigan BE. Role of oxidants in microbial pathophysiology. *Clin Microbiol Rev* 1997; 10: 1–18.
- [3] Sivakrishnan S, Muthu AK. In-vitro free radical scavenging activity of aerial parts of ethanolic extract of Albizia procera (Family: Mimosoideae). Int J Pharm Pharm Sci 2013; 5(2): 352–354.
- [4] Hamzah RU, Jigam AA, Makun HA, Egwim EC. Antioxidant properties of selected African vegetables, fruits and mushrooms: a review. In: Makun HA, editor. *Mycotoxin and food safety in developing countries*. Rijeka, Croatia: InTech; 2013.
- [5] Rahman A, Rahman MM, Sheik MMI, Rahman MM, Shadli SM, Alam MF. Free radical scavenging activity and phenolic content of *Cassia sophera L. Afr J Biotechnol* 2008; 7(10): 1591–1593.
- [6] Pal J, Ganguly S, Tahsin KS, Acharya K. In vitro free radical scavenging activity of wild edible mushroom, *Pleurotus* squarrosulus (Mont.) Singer. Indian J Exp Biol 2010; 48: 1210–1218.
- [7] Chang ST. Cultivation of Volvariella mushrooms in southeast Asia. In: Chang ST, Quimio TH, editors. Tropical mushrooms-biological nature and cultivation methods. Hong Kong, China: Chinese University Press; 1982.
- [8] Mattila P, Könkö K, Eurola M, Pihlava JM, Astola J, Vahteristo L, et al. Content of vitamins, mineral elements and some phenolic compounds in cultivated mushrooms. *J Agric Food Chem* 2001; 49: 2343–2348.
- [9] Aletor VA. Compositional studies on edible tropical specials of mushrooms. *Food Chem* 1995; 54: 256–268.
- [10] Fasidi IO. Studies on Volvariella esculenta (Mass) Singer: cultivation on agricultural wastes and proximate composition of stored mushrooms. Food Chem 1996; 55: 161–163.
- [11] Adebayo EA, Oloke JK, Ayandele AA, Adegunlola CO. Phytochemical, antioxidant and antimicrobial assay of mushroom metabolite from *Pleurotus pulmonarius*-LAU 09 (JF736658). J Microbiol Biotechnol Resour 2012; 2(2): 366-374.
- [12] Egwim EC, Ellen RC, Egwuche RU. Proximate composition, phytochemical screening and antioxidant activity of ten selected edible mushrooms. *Am J Food Nutr* 2011; 1(2): 89–94.
- [13] Keles A, Koca I, Genccelep H. Antioxidant properties of wild edible mushrooms. *Food Process Technol* 2011; doi:10.4172/2157– 7110.1000130.
- [14] Sofowora A. Medicinal plants and traditional medicines in Africa.2nd ed. Ibadan, Nigeria: Spectrum Books; 1993, p. 289.
- [15] Chang CC, Yang MH, Wen HM, Chern JC. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. J Food Drug Anal 2002; 10: 178–182.
- [16] Singleton VL, Orthofer R, Lamuela- Raventós RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods Enzymol* 1999; 299: 152-178.
- [17] Oloyede OI. Chemical profile of unripe pulp of *Carica pagaya*. *Pak J Nutr* 2005; 4: 379–381.
- [18] Association of Official Analytical Chemists. Official methods of analysis of the Association of Official Analytical Chemists. 14th ed. Maryland, USA: Association of Official Analytical Chemists; 1984, p. 188.
- [19] Gyamfi MA, Yonamine M, Aniya Y. Free-radical scavenging action of medicinal herbs from Ghana: *Thonningia sanguinea* on experimentally-induced liver injuries. *Gen Pharmacol* 1999; **32**: 661–667.

- [20] Oyaizu M. Studies on product of browning reaction prepared from glucosem mine. Jpn J Nutr 1986; 44: 307–315.
- [21] Ehssan HO, Saadabi AM. Screening of antimicrobial activity of wild mushrooms from Khartoum State of Sudan. *Microbiol J* 2012; 2: 64–69.
- [22] Wandati TW, Kenji GM, Onguso JM. Phytochemicals in edible wild mushrooms from selected areas in Kenya. J Food Res 2013; doi: 10.5539/jfr.v2n3p137.
- [23] Lee J, Lim S, Kang SM, Min S, Son K, Lee HS, et al. Saponin inhibits hepatitis C virus propagation by up-regulating suppressor of cytokine signaling 2. *PLoS One* 2012; doi: 10.1371/ journal.pone.0039366.
- [24] Thoppil RJ, Bishayee A. Terpenoids as potential chemopreventive and therapeutic agents in liver cancer cancer. World J Hepatol 2011; 3(9): 228–249.
- [25] Wang GY, Tang WP, Bidigare RR. Terpenoids as therapeutic drugs and pharmaceutical agents. In: Zhang LX, Demain AL, editors. *Natural products*. New Jersey, USA: Humana Press; 2005, p. 197–227.
- [26] Trease GE, Evans WC. Pharmacognosy: a physicians guide to herbal medicine. 13th ed. London, UK: Bailliere Tindall; 1989, p. 176–180.
- [27] Hamzah RU, Egwim EC, Kabiru AY, Muazu MB. Phytochemical and *in vitro* antioxidant properties of the methanolic extract of fruits of Blighia sapida, Vitellaria paradoxa and Vitex doniana. *Oxid Antioxid Med Sci* 2013; 2(3): 215-221
- [28] Nagavani V, Madhavi Y, Bhaskar RD, Koteswara RP, Raghava RT. Free radical scavenging activity and qualitative analysis of polyphenols by RP–HPLC in the flowers of *Couroupita guianensis* Abul. *Electron J Environ Agric Food Chem* 2010; 9(9): 1471–1484.
- [29] Parajuli S, Pun NT, Parajuli S, Jamakattel–Pandit N. Antioxidant activity, total phenol and flavonoid contents in some selected medicinal plants of Nepal. JHAS 2012; 2(1): 27–31.
- [30] Pereira DM, Valentão P, Pereira JA, Andrade PB. Phenolic: from chemistry to biology. *Molecules* 2009; 14: 2202–2211.
- [31] Jose GS, Radhamany PM. In vitro antioxidant activities, total phenolics and flavonoid of wild edible mushroom Macrolepiota mastoidea (Fr.) Singer. Int J Pharm Pharm Sci 2013; 5(2): 161–166.
- [32] Abugri DA, McElhenney WH. Extraction of total phenolic and flavonoids from edible wild and cultivated mushrooms as affected by different solvents. *J Nat Prod Plant Resour* 2013; 3(3): 37–42.
- [33] Abdullah N, Ismail SM, Aminudin N, Shuib AS, Lau BF. Evaluation of selected culinary-medicinal mushrooms for antioxidant and ACE inhibitory activities. *Evid Based Complement Alternat Med* 2012; doi: 10.1155/2012/464238.
- [34] Nabavi SM, Ebrahimzadeh MA, Nabavi SF, Fazelian M, Eslami B. In vitro antioxidant and free radical scavenging activity of Diospyros lotus and Pyrus boissieriana growing in Iran. Pharmacogn Mag 2009; 4(18): 123-127.
- [35] Gulcin L, Sat GI, Beydemir S, Elmastas M, Kufrevioglu OI. Comparison of antioxidant activity of clove (*Eugenia caryophylata* Thumb.) buds and lavender (*Lavandula stoechas* L). Food Chem 2003; 87: 393-400.
- [36] Dastmalchi K, Dorman HJD, Laakso I, Hiltunen R. Chemical composition and antioxidative activity of Moldavian balm (*Dracocephalum moldavica* L.) extracts. *LWT-Food Sci Technol* 2007; 40(9): 1655-1663.