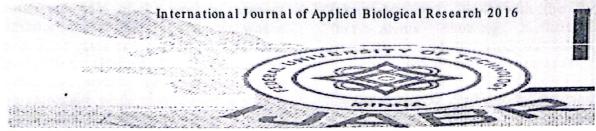
IJ ABR Vol. 7(2): xxx - xxx (2016)



Original Article

MOLLUSCICIDAL POTENTIALS OF Opuntia ficus-indica (CACTUS PLANT) ON Biomphalaria pfeifferi AND Bulinus globossus (MOLLUSCA: PLANORDIDAE)

Abdulazeez, I. L., Abolarinwa, S. O., Adeniyi, K. A*., Aliyu, R. H., Onoja, J. E. and Eke, S. S.

Entomology and Parasitology Unit, Department of Biological Sciences, Federal University of Technology, Minna, Nigeria.

Submitted: October, 2016; Accepted: November, 2016; Published: December, 2016

ABSTRACT

The use plant molluscicides have received considerable attention in the search for cheaper alternatives to chemotherapy and synthetic molluscicides in schistosomiasis control. Molluscicidal activities of Opuntia ficus indica (Cactus) extract was tested on Biomphalaria pfeifferi. and Bulinus globossus, the intermediate hosts of Schistosoma mansoni and Schistosoma haematobium, under standard laboratory co. The molluscicidal activity was assessed by determining the efficacy of various graded concentrations (0, 8, 16, 32, 64 and 128 mg/L) of the aqueous plant extracts on the two snail species. Ten (10) adults of uniform size were immersed in 6×3 (replicated thrice) trough. From the result, the highest molluscicidal activities were recorded at 128 mg/L on both snail species. B. pfefferthad the highest mean mortality 6.33 among both species at 24 hours exposure period. At 72 hours the mean mortality of both species were almost the same, however, the highest potency was recorded for B. pferiferri. The LC50 and LC90 of the O. ficus recorded on B. globossus were 12'37mg/L and 6.91mg/L respectively while LC50 and LC90 recorded on B. pferifferi were 5.28mg/L and 1.95mg/L respectively. The result obtained showed that Opuntia ficus-indica is a promising plant molluscicide candidate.

Keywords: Opuntia ficus-indica, Potency, Molluscicide, Extracts.

*Corresponding Author: adeniyikamoru.a@gmail.com

INTRODUCTION

Schistosomiasis is one of the most important public health problems after malaria (Muhammed et al., 2009). It's a

widespread disease infecting over 240 million people (Fenwick et al., 2006). It is found in tropical countries in Africa, Caribbean, Middle East, South America and South East Asia. However an

estimated 85% of the world's cases of schistosomiasis are in Africa, where prevalent rates can exceed 50% in local populations (Hamed, 2006). Out of the infected, over 120 million symptomatic while over 20 million have severe disease (Steinmann et al., 2006). A further 779 million are at risk of infection in tropical and subtropical areas of the world (Liu et al., 1997). Fresh water snails plays vital role in the transmission of schistosomiasis. Other factors include: lack of sufficiently trained health workers. inadequate safe water Supply, high cost of chemicals for snail control therapeutic drugs (Amita and Singh, 2001). Schistosomes, the causative agents of the disease have an indirect life cycle requiring aquatic snails, as intermediate host, for their transmission to man. Thus reduction of the parasite transmitting snails plays an important role among the strategies to control the disease (John 2004). Schistosomiasis can be controlled by destroying the carrier snail and thus breaking the life cycles of the parasite (WHO 2004). Snails of the genus Biomphalaria are the intermediate host for Schistosoma mansoni with the group pfeifferi being the most common in Nigeria.

Molluscicides are not frequently used as a major control mechanism due to cost implications and toxicity effects on nontarget organisms such as fish (WHO 2004; Adedeji and Okocha, 2012). Therefore there is need for search for a cheap, nontoxic and safer molluscicide (Molgaard et al., 1999; Mossoud and Habib, 2003). Consideration of cost and environmental effect of most molluscicides in current use for the control of schistosomiasis has generated the search for cheaper and less polluting molluscicides from natural source, especially those of plant origin

(Osuna-Martinez, 2014) with the hope that plants showing molluscicidal properties could be used on self-help basis to control diseases in rural areas.

Cactus (Opuntia ficus-indica) commonly known as prickly pear belongs to the family Cactaceae. Opuntia ficus indica produces sweet, nutritionally rich edible fruits; its tender cladodes are used as fresh green vegetable and salad (WHO 2004). The plant has been used in traditional folk medicine because of its role in treating a number of diseases and including conditions, diabetes, hypertension, hypercholesterolemic, rheumatic pain, gastric mucosa diseases and asthma, in many countries over the world (Reyes-Aguero 2005). Previous researchers have focused investigations for studying genus Opuntia in order to discover the properties of plant that could form the basis of their use in the prevention and cure of chronic diseases. Therefore clinical pharmacologic interest in the efficacy and safety of the phytochemicals present in genus Opuntia has grown during recent years due to the realization that many people self-medicate using this plant. The aim of this study is to evaluate the molluscicides potential of Opuntia ficus indica leaf extract on Biomphalaria pfeifferi and Bulinus globossus snails.

MATERIALS AND METHODS

Animal material

This study was conducted on Adult stages of Biomphalaria pfeifferi and Bulinus globosus snails of different diameter. In the laboratory, snails were identified to species level using identification keys by Otarigbo et al. (2013). Adults snails were maintained in separate plastic containers with capacity of about 1000ml of

of 10 snail/L at room temperature, (25-30°c). Green Lactuca sativa was immersed in boiling water for about one minute and then cool with cold water and were used to fed the snails in reasonable quantities at 2 days interval and water was changed twice a week, according to need. Dead snails were removed when observed.

Plant extraction

Opunia ficus indica whole plants were collected within the premises of Federal. University of Technology Minna. They were taken to the laboratory in a wet sack (to avoid direct exposure to sunlight which may lead to dehydration (Adetunji and Salawu, 2010). The plant was rinsed with distilled water to remove dust, sand, unwanted material and the healthy leaf was selected. Identification of the plant was confirmed by specialist in plant identification, in the department of Biological Sciences, Federal University of Technology, Minna, Niger state. The authentic plants were preserved in the herbarium section of Biological Science Laboratory. Three hundred (300) grams of the plant was pounded using mortar and pestle. The liquid extracts were squeezed using sieving net into a beaker. The extracts were concentrated under reduced pressure at 40atm using rotary evaporator.

Bioassay and Molluscicidal Potency Test of Adult Snail

The molluscicidal potency tests were carried out according to the standard method described by Otarigbo et al. (2013) and Adetunji and Salawu (2010). One gram (1g) of each crude extracts was dissolved in 10ml of distilled water and was later transferred into 490_ml of distilled water to prepare the stock solution (500ml). The different volumes

of 0.0 (control), 8, 16, 32, 64 and 128mL from the stock solution of the aqueous extract of the plant were added to an equal volume (500 ml) of dechlorinated water in plastic troughs (10 cm depth × 17 cm in diameter), to have working solutions. Then the concentration of each solution was calculated in milligram per Litre (mg/L): 0.0, 16, 32, 64, 128, and 256 mg/L. Ten (10) adults of uniform size were immersed in 6×3 replicates. In each set up, the snails were prevented from crawling out of the troughs by means of a fine mesh white cloth used for cover and tied to trough by rubber band. The snails were not fed during the course of the experiment; it had been observed that healthy snails live up to five days or more without food (Swaleh and Salawu, 2010), provided other environmental conditions are constant. After 24 hours exposure to the different plant extract concentrations. the snails were transferred to fresh dechlorinated water and maintained there for another 24 hrs. Molluscicidal test with the plant extract doses were separately repeated twice. Death of the snails was determined and confirmed by the lack of reaction to irritation of the foot with a blunt wooden probe to elicit typical withdrawal movements, mortality counts were recorded.

Data Analysis

Descriptive statistical analysis was performed using mean±standard error of mean. The results relating to molluscicidal activities of the plant extract and tested concentrations were evaluated using Analysis of Variance and Duncan Multiple Range Test, with a significance level of α =0.05. The median and Upper lethal concentrations were determined by regression analysis. All the analysis was carried out using Statistical Packages for Social Sciences (SPSS) 20th version and

Microsoft excel 2010.

RESULTS

The Molluscicidal potency of O. ficusindica on B. globosus snail species is presented in Table 1. The plant extract showed a significant mortality as the time of exposure moved from 24 hrs to 72 hrs. At 24 hrs exposure period, the mortality. recorded was 8 mg/L (1.00±0.00). There were no significant difference in the mortality recorded for 16 mg/L, 32mg/L and 64mg/L at 24 and 48 hrs exposure period. The mortality recorded for all concentration was significantly higher (P<0.05) than that of control. This was same after 48 hrs exposure period. At 48 control $(0.67\pm0.07),$ 8mg/L hours (2.33 ± 0.33) and 32mg/L (3.33 ± 0.88) recorded no significance different in snail mortality (P>0.05). At 72 hours exposure period, the highest effect on snail was recorded at 128 mg/L (8.33±0.33), however, this was not significantly difference from snail mortality recorded for 64 mg/L(7.67 ± 0.33).

The results on Table 2 showed the Mean Mortality of B. pfeifferi Adult Eexposed to Opuntia ficu- indica exract over a period of 72 hours. At 24 hours, control (0.33±0.03) snail mortality was not significantly different from 8 mg/L at p > 0.05, 128 (1.33 ± 0.67) (6.33±0.33) had the highest mollusc mean mortality and was significantly different from all other concentration at p<0.05. Also at 48 hours 128 mg/L (8.33±0.33) recorded the highest snail morality and was significantly different from all other mortality recorded at . 48 hrs exposure period.

The medial (LC₅₀) and upper (LC₉₀) lethal

concentration of the effect of O. ficus extract was presented in Table 3_The LC₅₀ and LC₉₀ of O. ficus against B. globusus and B. pfeipfferi were (6.91, 12.37) and (1.95, 5.28) respectively. The R² values recorded confirm the dependence of the mortality observed to the increment in the plant extract

Table 1: Mean Mortality of B. globosus Adult snail Exposed to Opuntia ficus indica Exract

ing stranging months	Hours			
Concentration (mg/L)	24	48	72	
CONTRO <u>L(0)</u>	0.00±0.00ª	0.67±0.17ª	0.67±0.07ª	_
8	1.00±0.00b	2.33±0.33*	4.67±0.67°	
16	1.33±0.33b	3.33±0.88ªb	3.67±0.88b	
32	1.67±0.33b	4.67±0.88b	5.67±0.33 b	
64	2.67±0.33b	5.67±0.88b	7.67±0.33°	
128	4.00±0.58°	6.33±0.33b	8.33±0.33°	

Values with the same superscript in the same column are not significantly different at p>0.05 Values are presented in mean±standard error of mean of two determinations

Table 2: Mean Mortality of Biomphalariam pferifferi Adult snail Exposed on Opuntia ficus-indica Exract

Concentration (mg/L)	24	48	72
Control(0)	0.33±0.03ª	0.67±0.17 °	0.67±0.58 a
8	1.33±0.67*	2.67±0.88ab	4.00±0.58b
16	2.33±0.33b	4.33±0.88b	5.00±1.00b
32	3.00±0.00 b	4.33±0.88b	6.00±0.58bc
64	3.67±0.33b	5.33±0.88b	7.67±0.88°
128	6.33±0.33°	8.33±0.33°	8.67±0.33°

Values with the same superscript in the same column are not significantly different at p>0.05 Values are presented in mean±standard error of mean of two determinations

Table 3: Léthal Concentrations of the Opuntia ficus-indica Extract on Adult snails

Snail species	LC90	LC50	R ²	Regression Equation
B. globosus	1237	6.91	0.909	Y=7.333 x-0.666
B. pfeipfferi	5.28	1.95	0.999	Y=12x+26.66

DISCUSSION

The environmental and economic problems associated with the use of synthetic molluscicides encouraged the search for substitutes of plant origin. At present, increasing attention is currently · given to the study of plant molluscides in hope that they may prove less toxic, costeffective, readily available and easily applicable by simple technique. So, the present_study is intended to search for i ideal molluscicide of plant origin on the basis of dosage response for combating adults snail intermediate host Schistosemiasis.

From the established regression lines of the tested extract, the molluscicidal activity was similar to that illustrated by Ebenso (2004) and Iglesias et al. (2008), against terrestrial land snail; Ederly et al. (1998), against aquatic snails including B. alexandrina, this could be as a result of slow stimulus response to the extract as indicated in their research. Furthermore, the toxicity of the tested snails has been . shown to increase as the concentration increases. The observed sensitivity of adult towards higher concentration is consistent with Stednichenko (2010) and Kirichuk et al. (2010). As reported by several investigators, elevated plant concentrations have extract been associated with an aggravated toxic response for several aquatic species-.

The susceptibility of Biomphalaria to the plant extract when compared to Bulinus is in agreement with the work of Munnert et al. (2003), As reported by many investigators (Abdel-kader and Sheraf, 2000; Abdel- Hamid, 2003), many invertebrate appear to be more tolerant to some substance. From the present

study, it would appear that extract bioaccumulation could be particular in the longevity of Bulinus. Some species, however, had a much greater tendency to accumulate one toxicant than the other (Munnert et al., 2003). Such differences are mainly related to the mode of exposure, rate of adsorption, species differences and the physiological and histopathological effects induced by the tested metals (Sheraf, 2000).

In the present study, the molluscicidal potency of the plant extract on the longevity of B. pfeifferi was also evaluated. From the established regression lines, the molluscicidal activity based on LC, values falls within the range of other plants that have Judged as promising been molluscicides (Abdel-kader and Sheraf. 2000; Abdel- Hamid, 2003; De et al. 2005). The recorded toxic effects on the longevity of the studied snails mainly attributed to several factors including plant specific differences of the extracted active ingredient, types of extracted products, differences in their mode of action, method of penetration and the behavioral characteristics of the studied animal (Mantawy et al., 2005). It is now well established that in many plants including the tested plants, the activity is due to the presence of bioactive phytochemicals including saponin contents (Rawi et al., 1996), tannins compounds (Bezera et al., 2002), triterpenoid and alkaloid components (Singh et al., 2010).

CONCLUSION

The study carried out showed that Opuntia has molluscicidal potential against Biomphalaria pfeifferiand Bulinus globossus snails which suggest the plant extract as an alternative option to synthetic molluscicides. Further studies are required to determine the mode-of-action of the plant extract and the damage they cause to the snail tissues as well as the side effects that may exist to the wildlife that surround them, through semi-field trials.

REFERENCES

Abdel-Hamid, H. (2003). Molluscicidal and in vitro schistosomicidal activities of the latex and some extracts of some plants belonging to Euphorbiacea. Journal of Egypt Society Parasitology, 33: 947-954.

Abdel-Kader, A. and Sharaf El-Din, A (2000). Potency of copper sulphate, bayluscide, Calendula micrantha officinalis and Ammi majus flowers on egg masses of B. alexandrina and Bulinus truncates. Egypt Journal of Schistosomiasis End Infectious Diseases, 22:159-173.

Adedeji O. B. and Okocha R. O. (2012). Overview of Pesticide Toxicity in Fish. Advances in Environmental Biology, 6(8): 2344-2351.

http://www.aensiweb.com/old/aeb/201 2/2344-2351.pdf

Adetunji, V. O. and Salawu, O. T. (2010). Efficacy of ethanolic leaf extracts of *Carica papaya* and *Terminalia catappa* as molluscicides against the snail intermediate hosts of schistosomiasis. *Journal Medicinal Plant Resources*, 4(22): 2348-2352.

Amrita, S., and Singh, N. K. (2001). Molluscicidal activities of the custard apple (Annona sqaunwsa I) alone and in combination with other plant derive molluscicide. *Herbs, Spices and Medicinal Plants* 8,31-36.

Bezerra, C., Silva, A., Ferreira, D., Ferri, P. and Santos S.(2002). Molluscicidal activity against *Biomphalaria glabrata* of Brazilian cerrado medicinal plants. *Fitotropia*, 73: 428-430.

Dallinger, R., Berger, C., Gruber, P., Hunziker, S. and Sturzenbaum, S. (2000). Metallothioneins in terrestrial invertebrates: Structural aspects, biological significance and implications for their use as biomarkers. Cellular Molecular Biology, 46: 331-346.

De, S. Luna, J., De, S. Santos, M., De Lim, M. C. De Omenta, F. and Sat'Ana, A. E. (2005). A study of the larvicidal and molluscicidal activities of some medicinal plants from northeast Brazil. Journal Ethnopharmacology, 97: 199-206.

Ebenso, I. E. (2004). Molluscicidal effects of neem (Azadirachta indica) extracts on edible tropical land snails. Pest Management Science, 60: 178-182.

Erdely, L., Kovacs, T. and Foldesi, Z. (1998). Pharmacological actions of essential (zinc, copper) and toxic metal ions (lead, mercury) on neurons and neuronal synapses of snail *Helix pomatia* L. *Neurotoxicology*, 19: 565 - 569.

Fenwick, A., Keiser, J., and Utzinger, J. (2006). Epidemiology, burden and control of schistosomiasis with particular consideration to past and current treatment trends. *Drugs Future*, (31): 413 - 425.

Hamed, M. A. (2010). Strategic control of

schistosome intermediate host. Epidemiolology, (3), 123 - 140.

Iglesias, J., Castillejo, J., Parama, R. and . Mascato, R. (2000). Susceptibility of the eggs of the pest slug *Deroceras reticulatum* to contact with metal salts. *Journal of Molluscan Studies* 66: 171 - 176.

John, O. (2004). Indigenous plants and schistosomiasis control in South Africa. Boletin Latinoamericano y del Caribe de Plantas Medicinales y Aromaticas 3, 8 - 22.

Liu, S. Y., Sporer, F., Wink, M., Jourdane, J., Henning, R., Li, Y. L. and Ruppel, A(1997). Anthraquinones in *Rheum palmatum* and Rumex dentatus (Polygonaceae), and phorbol esters in Jatropha curcas molluscicidal (Euphorbiaceae) with activity against the schistosome vector snails Oncomelania, Biomphalaria and Bulinus. Tropical Medicine and. International Health, 2:179 - 188.

Madsen, H., Simonsen, P. E., and Christensen N. Q. (1989). A basic field guide to the epidemiology of African Danish Schistosomiasis II. human WHO ' Bilharziasis Laboratory. Applied Center for Collaborating Malacology and Schistosomiasis Control, Charlottenlund, Denmark.

Mantawy, M., Hamed, E., Sammour, E. and Sanad, E.(2004). Influence of *Capparis spinosa* and *Acacia arabica* on certain biochemical haemolymph parameters of *Biomphalaria alexandrina Journal Egypt Social Parasitology*, 34:659-677.

Massoud, A. M. and Habib, F. S. (2003). The effects of Myrrh *Commiphora molmol* on the infected snails of *Schistosoma* species and their egg masses: effect on

shedding of cercariae and on snail fecundity. *Parasitology*, 33: 585-596.

Mohammed, Y. A. (2009). Studies on Molluscicidal Activity of Some Plants from Darfur against *B. truncatus* with Emphasis on *Alternanthera nodiflora* (Amaranthaceae) Phd thesis, University of Khartoum pp 86.

Molgaard, P., Chihaka, A., Furu, P., Ndekha, A. and Nyazema, N. (1999). Community based schistosomiasis control in Zimbabwe using the molluscicidal berries of *Phytolacca dodencadra* AETFAT conference in Harare, Zimbabwe.

Mummert, A., Neves, R., Newcomb, T. and Cherry, D. (2003). Sensitivity of juvenile freshwater mussels (*Lampsilis fasciola*, *Villosa iris*) to the total and un-ionized ammonia. *Environmental Toxicology Chemicals*, 22: 2545 - 2553.

Osuna-Martínez, U., Reyes-Esparza, J. and Rodríguez-Fragoso, L.(2014). Cactus (*Opuntia ficus-indica*): A Review on its Antioxidants Properties and Potential Pharmacological Use in Chronic Diseases. *Natural* Products Chemistry & *Research*, 2: 153

Otarigho, B. and Olajumoke, A.M.(2013). Molluscidal effect of aqueous and ethanolic extracts of Lemon grass (Cymbopogen citrallus) leaf against the different developmental stages of Biomphalaria pfeifferi. New York Science Journal, 5 (8): 70-77

Rawi, S., El-Gindy, H. and Abdel-Kader, A (1996). New possible molluscicide from *Calendula mircantha* officinalis and *Ammi majus*. II. Molluscicidal, physiological and egg laying effects against *Biomphalaria alexandrina* and *Bulinus truncates*.

Reyes-Agüero, J.A., Aguirre-Rivera, J.R. and Hernandez, M.H. (2005) Systematic notes and detailed description of *Opuntia ficus-indica* (L.) MILL. (Cactacea). *Agrociencia*, 39:395-408.

Singh, A. and Singh, V. (2009). Molluscicidal activity of *Saraca asoca* and *Thuja orientalis* against the fresh water snail *Lymnaea acuminata. Vetenary Parasitology*, 164: 206-210.

Singh, K., Yadav, P. and Singh A. (2010). Molluscicides from some common medicinal plants of Eastern Uttar Pradesh, India. *Journal of Applied Toxicology*, 30:1-7.

Stadnichenko, A and Kirichuk, G. (2000). The effect of ammonium nitrate on the residual nitrogen content in the hemolymph of pulmonate snail *Planorbarius purpura* (Mollusca: Pulmonata: Bulinidae) normally and in trematode invasion. *Parazitologiia*, 34: 402-407.

Steinmann, P., Keiser, J., Bos, R., Tanner, M. and Utzinger, J. (2006). Schistosomiasis and water resources development: systematic review, meta-analysis, and estimates of people at risk. *Infectious Diseases* 6: 411 - 425.

Swaileh, K. M. and Ezzughayyar, A. (2000). Effect of dietary Cd and co. on feeding and growth rate of the landsnail *Helix engaddensis*. *Ecotoxicology and Environmental Safety*, 47: 253 - 260.

World Health Organization. (2004). Prevention and control of schistosomiasis and soil-transmitted helminthiasis. Geneva: WHO Technical Report Series No.